

AD\_\_\_\_\_

Award Number: 95MM5571

TITLE: Cardiovascular Control of and Responses to Vasoconstrictor Hormones During Hypoxia

PRINCIPAL INVESTIGATOR: John R. Claybaugh, Ph.D.

CONTRACTING ORGANIZATION: Tripler Army Medical Center  
Tripler Army Medical Center, Hawaii 96859-5000

REPORT DATE: December 1999

TYPE OF REPORT: Final

PREPARED FOR: U.S. Army Medical Research and Materiel Command  
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for public release;  
distribution unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

DTIC QUALITY INSPECTED 3

20000428 133

# REPORT DOCUMENTATION PAGE

*Form Approved  
OMB No. 074-0188*

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503.

1. AGENCY USE ONLY (Leave blank)	2. REPORT DATE	3. REPORT TYPE AND DATES COVERED	
	December 1999	Final (1 Feb 95 - 31 Dec 99)	
4. TITLE AND SUBTITLE Cardiovascular Control of and Responses to Vasoconstrictor Hormones During Hypoxia			5. FUNDING NUMBERS 95MM5571
6. AUTHOR(S) John R. Claybaugh, Ph.D.			
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Tripler Army Medical Center Tripler Army Medical Center, Hawaii 96859-5000		8. PERFORMING ORGANIZATION REPORT NUMBER	
E-MAIL: john.claybaugh@amedd.army.mil			
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012		10. SPONSORING / MONITORING AGENCY REPORT NUMBER	
11. SUPPLEMENTARY NOTES			
12a. DISTRIBUTION / AVAILABILITY STATEMENT Approved for public release; distribution unlimited			12b. DISTRIBUTION CODE
13. ABSTRACT ( <i>Maximum 200 Words</i> ) These studies addressed the role of certain factors on the ability to maintain blood pressure (BP), cardiac output (CO) and O <sub>2</sub> delivery (O <sub>2</sub> del) during hemorrhage in the conscious goat. We studied the effects of oxygen content of inspired air, the presence of the spleen, roles of arginine vasopressin (AVP), the renin-angiotensin system, and the presence of estrogen. We observed that a controlled hemorrhage at 0.5 ml/kg/min for 30 min conducted in the same goats while exposed to either 11, 21, and 100% FIO <sub>2</sub> , reduced mean arterial BP by approximately 25, 15, and 5 mmHg respectively. Improved maintenance of BP during hyperoxia was achieved by an earlier rise in systemic vascular resistance, and O <sub>2</sub> consumption was similar in all experiments following hemorrhage. Presence of the spleen did not affect the magnitude of drop in BP, O <sub>2</sub> del, nor hormonal responses to the hemorrhage. In other experiments, i.v. infusions of AVP in goats breathing 11% FIO <sub>2</sub> increased the arterial O <sub>2</sub> concentration, but the AVP-induced decrease in cardiac output prevented an improvement in O <sub>2</sub> del. Neither hypoxia nor the rate of AVP infusion affected whole body AVP clearance. Lastly, we observed that estrogen administration to gonadectomized female or male goats lowered the hypotensive threshold during hemorrhage because of a reduced ability to maintain CO.			
14. SUBJECT TERMS Hemorrhage, Cardiovascular, Splenectomy Oxygen Delivery, Oxygen Consumption, Hypoxia, Hyperoxia, Vasopressin, Renin, Estrogen			15. NUMBER OF PAGES 16
16. PRICE CODE			
17. SECURITY CLASSIFICATION OF REPORT Unclassified	18. SECURITY CLASSIFICATION OF THIS PAGE Unclassified	19. SECURITY CLASSIFICATION OF ABSTRACT Unclassified	20. LIMITATION OF ABSTRACT Unlimited

NSN 7540-01-280-5500

Standard Form 298 (Rev. 2-89)

Prescribed by ANSI Std. Z39-18  
298-102

FOREWORD

Opinions, interpretations, conclusions and recommendations are those of the author and are not necessarily endorsed by the U.S. Army.

*JL* Where copyrighted material is quoted, permission has been obtained to use such material.

*JL* Where material from documents designated for limited distribution is quoted, permission has been obtained to use the material.

*JL* Citations of commercial organizations and trade names in this report do not constitute an official Department of Army endorsement or approval of the products or services of these organizations.

X In conducting research using animals, the investigator(s) adhered to the "Guide for the Care and Use of Laboratory Animals," prepared by the Committee on Care and use of Laboratory Animals of the Institute of Laboratory Resources, national Research Council (NIH Publication No. 86-23, Revised 1985).

X For the protection of human subjects, the investigator(s) adhered to policies of applicable Federal Law 45 CFR 46.

X In conducting research utilizing recombinant DNA technology, the investigator(s) adhered to current guidelines promulgated by the National Institutes of Health.

X In the conduct of research utilizing recombinant DNA, the investigator(s) adhered to the NIH Guidelines for Research Involving Recombinant DNA Molecules.

N/A In the conduct of research involving hazardous organisms, the investigator(s) adhered to the CDC-NIH Guide for Biosafety in Microbiological and Biomedical Laboratories.

  
J.R. Clegg  
PI - Signature

22 Feb. 2000  
Date

## Table of Contents

	<b>Page</b>
<b>Cover.....</b>	<b>1</b>
<b>SF 298.....</b>	<b>2</b>
<b>Foreword.....</b>	<b>3</b>
<b>Introduction.....</b>	<b>5</b>
<b>Body.....</b>	<b>7</b>
<b>Key Research Accomplishments.....</b>	<b>9</b>
<b>Reportable Outcomes.....</b>	<b>14</b>
<b>Conclusions.....</b>	<b>15</b>
<b>References.....</b>	<b>15</b>

## INTRODUCTION

When the tissues of the body receive too little O<sub>2</sub> to maintain adequate function, various reflexes respond causing increased respiration and cardiac output. Depending on the cause of the reduced O<sub>2</sub> the systemic vasculature can constrict or dilate. In response to hypoxia, for instance, cardiac output is increased and systemic vascular resistance is reduced (1). This allows for increased flow of blood to the tissues, improved O<sub>2</sub> delivery, and maintenance of normal blood pressure. In the case of hemorrhage, however, the body is also faced with reduced O<sub>2</sub> at the tissues, but by reflexes mediated primarily via low pressure system baroreceptors, an increased sympathetic tone results leading to vascular constriction and increased heart rate (2). This allows the system to maintain blood pressure with hemorrhages up to about 15% of total blood volume, thereby maintaining perfusion pressure and circulation to select areas of the body. Near this volume of hemorrhage, however, there is a sympathetic inhibition. This appears to initiate the life threatening conditions of decreasing mean arterial blood pressure and heart rate with any further loss of blood.

It can be hypothesized from this background, that if hemorrhage were to occur during hypoxia that there would be a conflict of reflexes, hypoxia causing a vasodilatation and hemorrhage causing a vasoconstriction. In fact, we reported an earlier reduction in blood pressure during a controlled hemorrhage in goats during exposure to hypoxia, 10 % inspired O<sub>2</sub>, than when breathing a normoxic gas mixture (3). This was subsequently confirmed in experiments using rats that were hemorrhaged either during acute exposures to hypoxia (12% O<sub>2</sub>) or after a chronic, 3-week exposures to hypoxia (4). Only the acute exposure to hypoxia was associated with the decreased ability to maintain blood pressure during hemorrhage. Moreover, the chronic exposures increased the threshold of volume of blood loss required to elicit a fall in blood pressure in both the normoxic and hypoxic states (4,5). The mechanism of the latter is unknown.

It could also be hypothesized that if hypoxia causes a decreased ability to maintain blood pressure, that hyperoxia may reduce or eliminate the reduction in mean arterial blood pressure in response to a modest hemorrhage. However, the vasoconstrictor effects of hyperoxia may adversely affect oxygen delivery to certain areas of the body. Therefore, as one objective of these studies, we investigated and compared the ability to maintain arterial blood pressure during hemorrhage under conditions of normoxia, hyperoxia and hypoxia. In addition, measurements of systemic vascular resistance, O<sub>2</sub> delivery and O<sub>2</sub> consumption were determined.

A second objective of this proposal was to further investigate the effects of vasopressin on O<sub>2</sub> delivery and O<sub>2</sub> consumption. It has been known for many years that vasopressin can be released by stimulation of chemoreceptors (6). Furthermore, hypoxic stimulation of vasopressin appears very early in development, and has been reported in fetal sheep during the third trimester (7). While there exists strong evidence for a role of vasopressin as causal in the often fatal condition of high altitude cerebral edema, and possibly high altitude pulmonary edema (8), there has been a paucity of information concerning the possible beneficial role of vasopressin in the response to hypoxia. Intuitively, it would seem that such an early, ontogenetically, developed response, that is present in many if not all mammals, would have some survival value. In previous studies in our laboratory (9,10) we have shown that exogenous, intravenously administered, vasopressin greatly improves PaO<sub>2</sub>, SaO<sub>2</sub>, and shunt fraction in the anesthetized, artificially ventilated, new born piglet, especially during hypoxic conditions. This response has

not been confirmed in a conscious, spontaneously breathing animal. The adult goat model presently being used in our laboratory was very suitable for these experiments, because, in conjunction with local anesthesia, the jugular vein is readily accessible introduction of a Swan-Ganz catheter for the necessary measurements. In addition, the multiple infusion rates of vasopressin and subsequent plasma levels of vasopressin achieved allowed an opportunity to further study the whole body clearance of vasopressin. It has been previously shown to be positively correlated with plasma levels of vasopressin in dogs and humans (11). This is the first study to determine the effects of hypoxia on the whole body clearance of vasopressin.

The role of the spleen in animals with contractile spleens, has been thought to be that of a reservoir of blood. Therefore, in studies involving hemorrhage in animals with contractile spleens, the spleen is often removed, presumably so the animal will respond more like a human, because humans do not have contractile spleens (12). The spleen, however, may have other effects. For instance, Horton et al. (13) have reported evidence that dogs with their spleens intact, respond to hemorrhage with greater myocardial contractility than splenectomized dogs with or without augmented infusion of blood to mimic the effects of the spleen. Similar studies have not been done in the goat, nor on the modest rate and magnitude of hemorrhage that we have employed in this conscious standing animal. Therefore, in order to better understand the role of the spleen in our animal model we investigated the cardiovascular responses and the O<sub>2</sub> delivery and consumption and hormonal responses in the intact and splenectomized goat.

The third major objective of the proposal was directed at central control mechanisms that affect the maintenance of blood pressure and cardiac output during hemorrhage. Specifically we attempted to test the hypothesis that central mechanisms that are responsible for the sympathetic withdrawal are mediated either directly or influenced by vasopressin. These experiments were not completed due to technical difficulties and slow progress. Also, since the writing of this proposal other work has been reported that supports our hypothesis, and lessens the necessity to complete these studies as originally conceived. For instance, Budzikowski et al. (14) have reported that blockade of central V1 receptors for vasopressin abolished the hemorrhage-induced bradycardia and hypotension in WKY (Wistar Kyoto) rats, but not in SHR (Spontaneously Hypertensive Rats).

As a result of our slow progress and the recent work of others cited in the previous paragraph, we requested, and were granted permission, to postpone those studies and add a new objective during the last 18 months of the grant. Specifically, we have tested the hypothesis that estrogen will reduce the tolerance to hemorrhage. This hypothesis was based on the previous studies that demonstrated that the presence of estrogen reduced the pressor effects of infused angiotensin in sheep (15), and the pressor responsiveness to vasopressin is reduced in the female rat (16), and also, studies by Crofton et al (17) provided suggestive evidence that hemorrhage in female rats during proestrus (periods of high estrogen) resulted in an earlier onset of hypotension than similarly hemorrhaged female rats during other phases of the estrus cycle or than male rats. These later studies were based on the statistical significance of one time period at 10 minutes after the removal of 10% of the blood volume. The present studies were conducted to further these observations by determining if the effects may be due to gonadal steroids, specifically, estradiol-17 $\beta$ , whether the effects are due to reductions in CO or SVR, and whether the effects of estrogen can be manifest in both male and female animals after gonadectomy.

## BODY:

### *Experimental Methods:*

Young adult female and some castrated male goats, weighing about 30-45 kg, were used. This grant involved three protocols that were approved by the Tripler Army Medical Center Institutional Animal Care and Use and Committee. The facility and program are AAALAC approved.

All goats were prepared with an exteriorized carotid arterial loop, as described earlier (3), at least 3 weeks before any experiments. This allows repeated access to the artery for purposes of arterial pressure recordings, heart rate, arterial blood gas and cooximetry measurements, hormone samples, and blood removal for hemorrhage.

On the experiment day, the goats were placed in a stanchion and the areas of the carotid loop and the contralateral jugular vein were shaved and swabbed with antiseptic. The carotid artery was cannulated and the line connected to a pressure transducer and recorder. After local anesthesia, a small skin incision was made over the jugular vein, and an introducer was inserted into the jugular vein, and a Swan-Ganz catheter was inserted through the introducer into the jugular vein and advanced to the pulmonary artery. Location was determined by monitoring the pressures at the tip and at the point used to measure central venous pressure (CVP), while advancing the catheter. With these catheters in place, we measured mean arterial pressure (MABP), and the systolic and diastolic pressures, heart rate, CVP, pulmonary arterial pressure (PAP), pulmonary arterial wedge pressure (PAWP), and cardiac output (CO) was determined by thermodilution.

Infusions were made via the port to the area of the right atrium in the Swan-Ganz catheter, and blood removal was from the arterial or the Swan-Ganz catheter depending upon whether arterial or venous blood was needed. With assessments of O<sub>2</sub> content by cooximetry and blood gas analysis on the arterial and venous sites, and having measured cardiac out put, calculated values of O<sub>2</sub> delivery and O<sub>2</sub> consumption and of systemic vascular resistance and pulmonary arterial resistance were obtained.

In experiments where hypoxic or hyperoxic gas mixtures were administered, either air, air diluted with nitrogen (hypoxia), or 100% O<sub>2</sub> was delivered through a face mask through a one-way valve, from compressed gas cylinders. The fractional inspired O<sub>2</sub> (FIO<sub>2</sub>) was constantly monitored and flow meters were adjusted to deliver either 11%, 21%, or 100% O<sub>2</sub>.

Hemorrhage was performed by removal of blood with a peristaltic pump calibrated to remove 0.5 ml/kg/min. The blood was collected in sterile blood donor bags containing citrate. At the conclusion of the experiment the blood was returned to the animals.

Splenectomy was performed via a left paracostal approach originally described for splenectomy in sheep (18). Bilateral ovariectomy was performed via a single ventral midline incision, also as done previously in sheep (15).

### *Experimental designs:*

Hyperoxia series: Six goats were hemorrhaged 0.5 ml/kg/min while exposed to normal air, 11% O<sub>2</sub> or, 100% O<sub>2</sub>. Control measurements were obtained 60 minutes after cannulation, and then the inspired gas mixture was begun. After 30 min of breathing the prescribed gas mixture, the pre-

hemorrhage measurements were obtained. Hemorrhage was then begun, with measurements obtained at 10 min intervals up to 30 min. At this time the experiment was concluded and the hemorrhaged blood was returned.

Arginine vasopressin (AVP) infusion series: Six goats were infused with AVP at various doses while breathing either 11% or 21% O<sub>2</sub>. A set of measurements was obtained 60 minutes after cannulation. The breathing gas was then initiated, and normal saline (vehicle) was infused for 20 min and the second set of measurements was obtained. The third through the seventh measurements were obtained following 20 min periods of AVP infusion at 30, 100, 300, 1000, 3000 pg/kg/min in that order.

Splenectomy series: Six goats had blood volumes assessed by the Evan's Blue dye dilution technique before and after splenectomy. They also were hemorrhaged before and after splenectomy similar to the protocol described above in the "Hyperoxia series". Following the hemorrhage experiments in the spleen-intact condition, the surgery was performed and the second hemorrhage was conducted a minimum of 3 weeks later, and within 2 months of the first experiment.

Estrogen series: Six female goats were splenectomized and bilaterally ovariectomized. Three neutered male goats were also used. Two sets of experiments were conducted on all goats with or without preceding daily subcutaneous injections of estradiol-17 $\beta$  (100  $\mu$ g/kg). In the first set of experiments, conducted either without previous estrogen or following 2 weeks of daily estrogen, the goats were hemorrhaged at a rate 0.5 ml·kg<sup>-1</sup>·min<sup>-1</sup>. Prior to the hemorrhage and at 10 min, 20 min, and 30min or any period prior to 30 min when mean arterial blood pressure had fallen to 65 mmHg, the various cardiovascular parameters and blood samples were obtained. AVP, renin activity, and nor-epinephrine, hematocrit and osmolality measurements were obtained. Blood volume (Evan's Blue) was determined prior to each hemorrhage experiment in order to accurately determine the % blood volume loss required to lower blood pressure in the two states of circulating estrogen. Also, estradiol-17 $\beta$  was measured in the first blood sample to verify the effectiveness of the estrogen administration. In the second set of experiments the goats were infused with AVP (0.3, 10.0, 28.3 ng·kg<sup>-1</sup>·min<sup>-1</sup>), angiotensin II (1.25, 5, and 10 ng·kg<sup>-1</sup>·min<sup>-1</sup>), and phenylephrine (1.0, 1.8, and 6  $\mu$ g·kg<sup>-1</sup>·min<sup>-1</sup>), each dose for 10 minutes with a one-to two-hour recovery in between different hormone infusions. The order of the hormone infusions was randomized. During each infusion, measurements of arterial blood pressure (BP), CO, CVP, PAP, and PAWP were recorded, and stroke volume and systemic vascular resistance (SVR) calculated. These experiments were also conducted without the presence of estrogen and also after 21 days of daily estrogen injections.

Statistics: There was a minimum of six goats in each series. The same goats were in each exposure and were sampled over several periods. Therefore, the Analysis of Variance (ANOVA) design used was a one-way with repeated measures. That is, the repeated measures were obtained over three experiments (for the data in figure 1) with 5 measurements in each experiment. Therefore these data were analyzed as a 6 (goat) by 15 (treatments) table. When the F value for the ANOVA was significant, the means of interest were compared by post hoc

analysis with a Fisher's LSD Multiple-Comparison test. The statistical software used was NCSS version 6.0.21 (Copyright 1996, Dr. Jerry L. Hintze, Kaysville, Utah).

#### KEY RESEARCH ACCOMPLISHMENTS:

##### Hyperoxia series:

As hypothesized, the fall in mean arterial pressure in response to 30 min of hemorrhage at 0.5 ml/kg/min was dependent upon the concentration of O<sub>2</sub> in the inspired air (Figure 1).

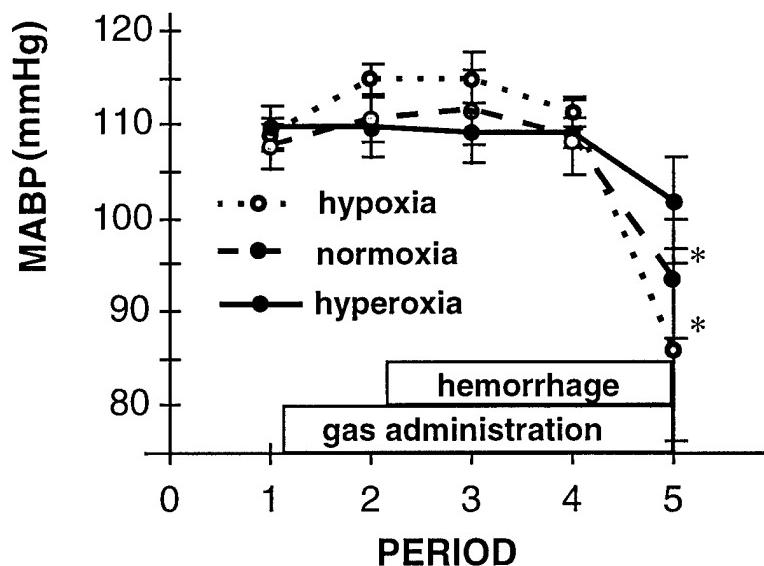


Figure 1. Response of mean arterial blood pressure to hemorrhage of 0.5 ml/kg/min. Period 1 = 60 min post cannulation, 2 = 30 min following initiation of either 11 %, 21% or 100% FIO<sub>2</sub>, 3 = 10 min of hemorrhage, 4 = 20 min hemorrhage, 5 = 30 min hemorrhage. \* = P < 0.05 compared to period 1.

It is noteworthy that with this magnitude of hemorrhage, 15/ml kg, inspiration of 100% O<sub>2</sub> prevented the fall in MABP, where both normoxic and hypoxic conditions were accompanied by significant decreases. Recent work by others appearing in an abstract has described a similar response in rats while breathing 100% O<sub>2</sub> following hemorrhage (19). Our work focused further on O<sub>2</sub> delivery and consumption. It was anticipated that 100% O<sub>2</sub> might improve the maintenance of arterial blood pressure, but at what consequence to O<sub>2</sub> distribution to the tissues? Decreased O<sub>2</sub> consumption (VO<sub>2</sub>) may be an index of poor perfusion of tissues or perhaps the reduced work involved in breathing. Regardless, VO<sub>2</sub> did decrease with the initiation of 100% O<sub>2</sub> breathing (Figure 2). However, during hemorrhage the VO<sub>2</sub> returned

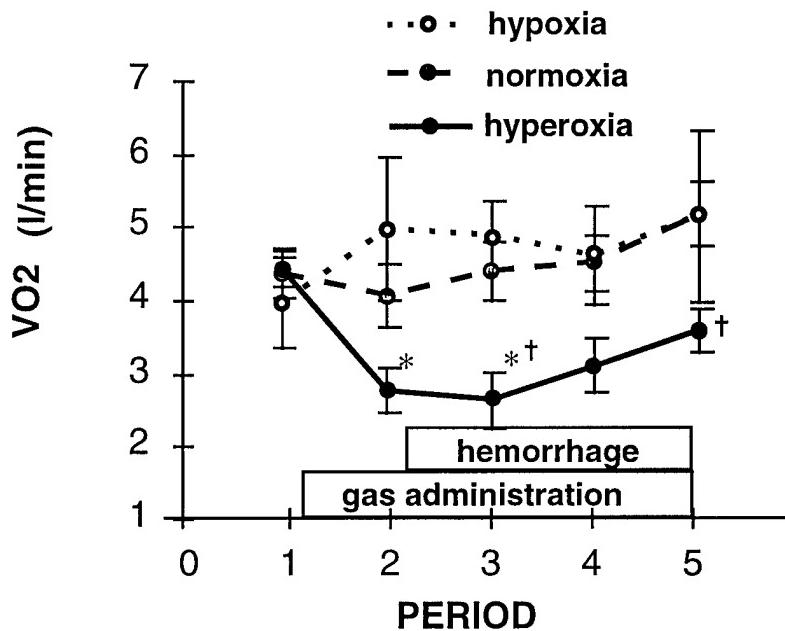


Figure 2. O<sub>2</sub> consumption (VO<sub>2</sub>) during hemorrhage in conscious goat while breathing 11%, 21% or 100% O<sub>2</sub>. The periods are the same as described for Fig.1., \* = P<0.05 compared to period 1, † = P < 0.05 compared to normoxic conditions.

toward baseline, but VO<sub>2</sub> was still significantly below the values observed while breathing hypoxic or normoxic gases. Since the gap in VO<sub>2</sub> between hyperoxic and normoxic gas breathing appears to remain constant as VO<sub>2</sub> increases during hemorrhage, it is tempting to speculate that the cause for the reduction does not change. If this is the case, then hyperoxia does not compromise the necessary circulation to provide O<sub>2</sub> to those areas of demand during hemorrhage. Therefore, administration of 100% O<sub>2</sub> may be a safe and physiologically useful adjunct to emergency first aide to patients during hemorrhage, because of its ability to maintain perfusion pressure. Regional blood flow distribution studies will be necessary to assess the validity of this speculation.

We also hypothesized that, due to stimulation of chemoreceptors by hypoxia, that the vasopressin response to hemorrhage might be altered. Vasopressin was increased during the last period in all situations, but was significantly blunted during hyperoxia. This is expected because of the differences in MABP (Fig 1). However, subsequent regression analysis between MABP and plasma AVP indicated similar slopes with all breathing gas mixtures. In contrast, the low pressure system, as measured by right atrial pressure, was also correlated with plasma AVP levels, and hyperoxia was shown to significantly reduce the plasma AVP response to reduced stretch of the low pressure system baroreceptors. Vasopressin alters the regional distribution of blood flow, greatly favoring blood flow to the brain and heart, while shunting it away from the kidney, spleen, muscle, and skin (20). Thus, the reduced vasopressin response during hemorrhage during hyperoxia may work against its beneficial effects.

Clearly more work is needed to sort out the beneficial from potentially harmful effects of 100% O<sub>2</sub> administration during hemorrhage, but our results favor a beneficial effect.

### Arginine vasopressin (AVP) infusion series:

The infusion of AVP into goats either while breathing hypoxic or normoxic gas mixtures resulted in nearly identical plasma levels of the hormone at all doses. Therefore, hypoxia resulted in no difference in AVP clearance contrary to observations made earlier on experiments in the neonatal piglet. However, exogenous administration of AVP did result in an increased PaO<sub>2</sub> (Figure 3). Note the significant improvement of PaO<sub>2</sub> at the highest infusion rate of AVP.

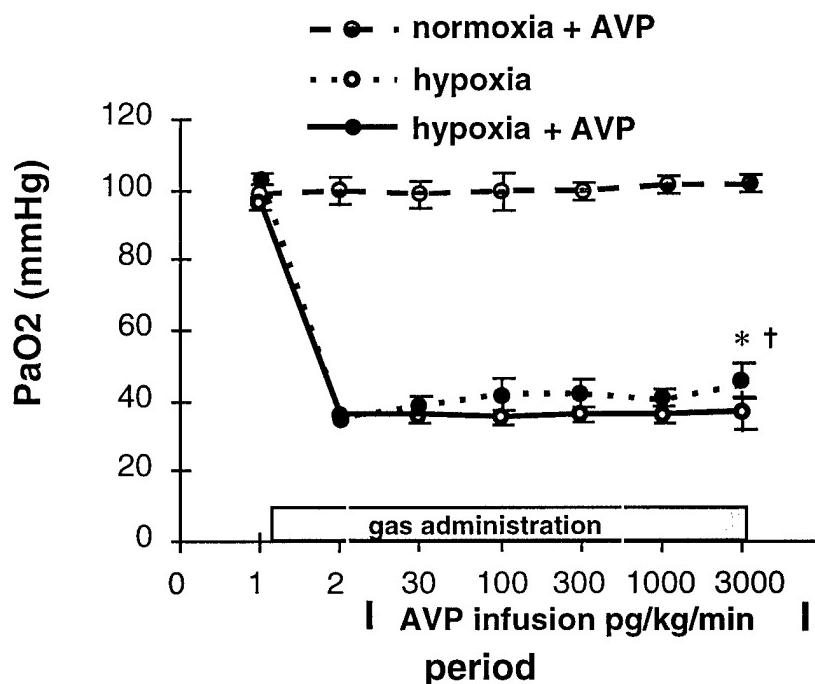


Figure 3. Effect of AVP infusion on PaO<sub>2</sub> during hypoxia and normoxia. Periods 1 and 2 are control periods 60 after catheterizations, and 30 min after initiation of 11% or 21% FIO<sub>2</sub>. AVP infusion rates are shown on the x axis, each infusion period was 20 min. \* = P<0.05 compared to period 2. † = P < 0.05 compared to hypoxia without AVP infusion.

This improved PaO<sub>2</sub> was in association with a 10% improvement in arterial hemoglobin saturation (P<0.05), but because of the effects of AVP in decreasing cardiac output, O<sub>2</sub> delivery was not significantly improved. Never-the-less the effects of AVP on PaO<sub>2</sub> were confirmed in this model, albeit a modest response. In previous work in the anesthetized piglet model, we were able to infuse LVP (lysine vasopressin) at much higher doses (100 ng/kg/min), but these doses would not be tolerated in the conscious adult goat. Thus, the dose may have contributed to more obvious response we observed in the piglet. The present experiments, however, demonstrate that this response can be achieved by physiological levels of plasma AVP. In these experiments the plasma AVP levels were approximately 135 µU/ml in both infusion experiments which is common during hypotensive hemorrhage.

### Splenectomy experiments:

Changes in MABP, cardiac output, and systemic vascular resistance were essentially identical when hemorrhage was performed before and after splenectomy. In addition, the hormone responses of vasopressin and renin were very similar. The most outstanding difference in the cardiovascular responses was the heart rate response shown below (Figure 4).

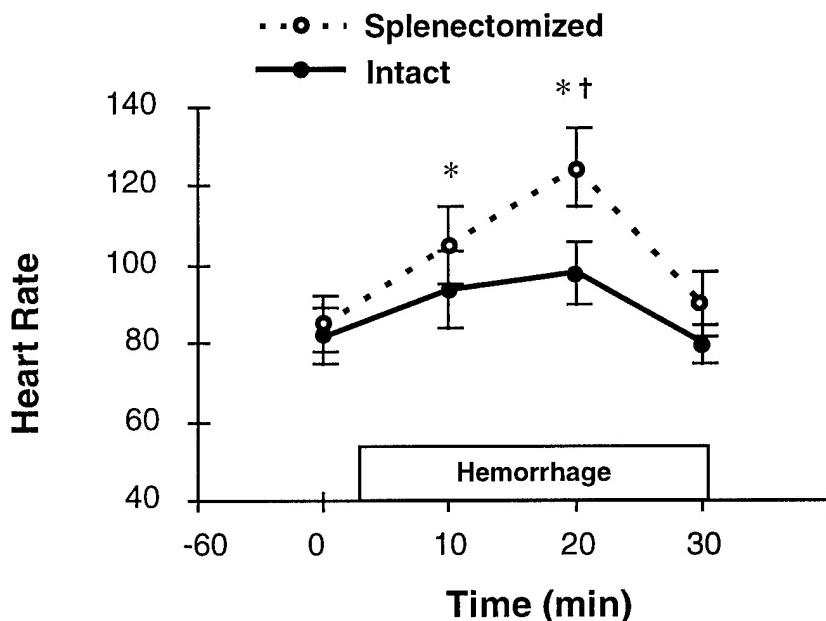


Figure 4. Effect of splenectomy on the heart rate response to hemorrhage (0.5 ml/kg/min). \* =  $P < 0.05$  compared to pre-hemorrhage value. † =  $P < 0.05$  compared intact condition.

Since the cardiac output was unchanged as a consequence of splenectomy, it would seem that stroke volume (SV) would have to be reduced in the splenectomized condition. In fact, mean values were lower, albeit not statistically significant, and this was due to a slightly reduced central venous pressure (CVP), also not significantly lower than in the intact state. Since hematocrit was significantly increased during the 20 and 30 minutes of hemorrhage, it would appear that the spleen was ejecting some volume of red blood cells into the circulation. This added volume may have contributed to the slightly improved CVP and SV during hemorrhage in the intact goat. Furthermore, it is likely that the inability to maintain SV in the splenectomized goat, reflexly resulted in increased sympathetic drive to maintain CO and adequate O<sub>2</sub> delivery. At this level of hemorrhage, in this model, the consequences of splenectomy on CO, O<sub>2</sub> delivery and consumption, and on hormonal responses to volume of hemorrhage was indiscernible.

### Estrogen administration experiments:

These experiments are not quite finished, and many of hormonal analyses remain to be performed. However, some data are clear enough to draw conclusions at this point. In Figure 5 it can be seen that, as we hypothesized, the amount of blood loss required to cause a drop in

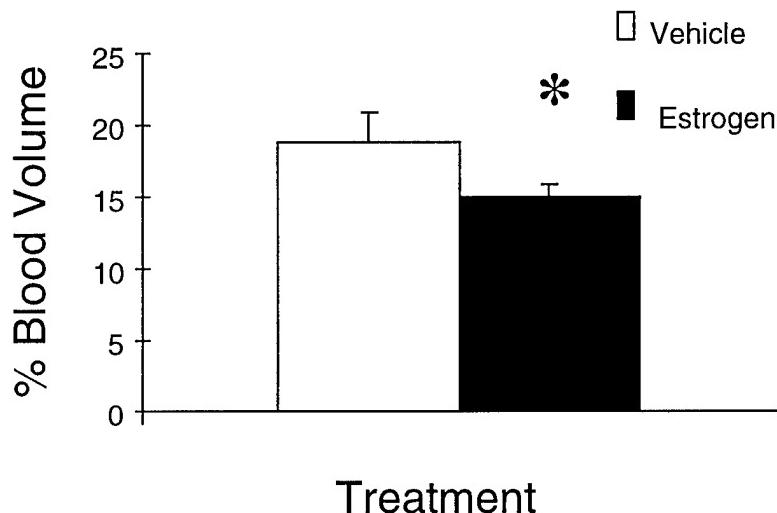


Figure 5. Effect of daily estradiol- $17\beta$  administration to ovariectomized female goats ( $n = 6$ ) on the blood volume loss required to induce a decrease in mean arterial pressure to 65 mmHg in response to hemorrhage (0.5 ml/kg/min). \* =  $P < 0.05$  compared to saline injected control state.

mean arterial blood pressure to 65 mmHg, is significantly reduced when the goats have been given daily estrogen treatment for 2 weeks. The serum estrogen levels achieved by the daily subcutaneous injections were approximately 200 pg/ml, similar to the estrogen levels reported by Magness et al. (15) in studies in the sheep where they infused estradiol- $17\beta$  intravenously for 2 weeks. Our data in 3 male goats revealed similar responses to the 6 female goats summarized in Fig 5. The apparent reason for this reduced tolerance to hemorrhage during estrogen administration, is not due to a reduced systemic vascular resistance as we had hypothesized, but rather a reduced cardiac output.

Furthermore, the data summarized in Figure 6 further support the lack of an effect of estrogen treatment on systemic vascular resistance. Note, even after 21 days of daily estrogen administration, the basal levels and hormone-stimulated values of systemic vascular resistance were not different between the control and the estrogen treated state. These findings are in contradiction to the findings of Magness et al. (15) and others, and we are as yet without a clear explanation. Several differences in the studies are evident; for example, the method of estrogen administration, the species, and the use of splenectomized animals in the present studies, but conjecture at this point is premature until all of the data has been analyzed.

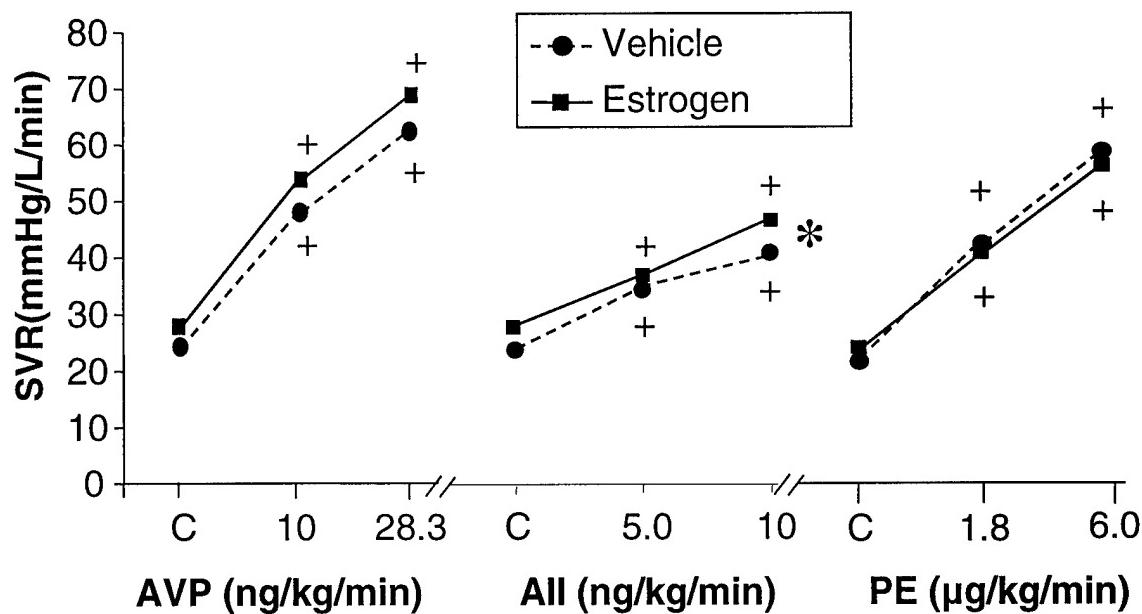


Figure 6. Effect of daily estradiol-17 $\beta$  administration for 3 weeks to ovariectomized female goats ( $n = 6$ ) on the responses of systemic vascular resistance (SVR) to vasopressin (AVP), angiotensin II (AII) and phenylephrine (PE). \* =  $P < 0.05$  compared to vehicle only at a similar time, + =  $P < 0.05$  compared to the control period (C) prior to hormone infusion.

#### REPORTABLE OUTCOMES:

Some of these studies have been presented in the following abstracts, and I anticipate that all of these will result in full manuscripts. In addition, the studies on the effects of estrogen on the hemorrhage response are nearly complete, but have not yet been presented, and these studies will result in a Ph.D.Thesis and they will be presented at the next Experimental Biology meetings in Orlando, Florida (2001). They too should result in a full manuscript.

Claybaugh, J.R., A.K. Sato, and C.F.T. Uyehara. Effects of vasopressin (VP) on the cardiopulmonary responses to hypoxia in the conscious, spontaneously breathing, goat. Experimental Biology 94. Abstract #4778, FASEB J. 8:A824, 1994.

Claybaugh, JR, Sato, AK, Van Scy, SC. Effects of Hyperoxia and hypoxia on cardiopulmonary and hormonal responses to hemorrhage. Experimental Biology '95, (Abstract # 1535), FASEB J. 9: A265, 1995.

Urada. K.K., Claybaugh, J.R., and Sato A.K., Splenectomy alters the normal response to a hemorrhage in the goat. Experimental Biology '97. New Orleans, LA. April 6-10, 1997. FASEB J.11:A462, 1997 (Abstract #2675).

## CONCLUSIONS:

The work in this project has produced the following findings: 1) Administration of 100 % O<sub>2</sub> during hemorrhage will help to maintain mean arterial blood pressure without compromising O<sub>2</sub> consumption. 2) Vasopressin improves PaO<sub>2</sub> during hypoxia, albeit very slightly, at physiological levels of plasma AVP. Also, hypoxia does not seem to alter the whole body clearance of AVP compared to the normoxic condition. 3) Splenectomy altered the responses to a slightly hypotensive hemorrhage in ways that lead to a clear difference in heart rate. Apparently owing to a reduced filling pressure, SV was not as well maintained as in the intact condition; however, CO was maintained by the increased heart rate. The mechanisms for this adjustment can not be determined from our data. 4) The presence of high levels of estrogen reduced the tolerance to hemorrhage in the conscious, splenectomized, ovariectomized goat.

Lastly, this grant has provided support to a graduate assistant, K.K. Urada for his pre-doctoral research, it provided considerable training experience for Dr. S.C. VanScoy, a neonatal fellow in our laboratory for 3 years. Mr Urada dedicated full time to the project for this past year with no stipend support for the last series of experiments that will be used for his Ph.D. Thesis to be defended at the end of summer. At the writing of this report there remain only two more experiments to be successfully executed before completion of the project. Manuscripts for the three completed studies are in various stages of preparation. The work developed in reference citation (3), and in the first abstract above, have resulted in an invitation for a Symposium on Sports Science at the '97 Winter Olympics in Nagano Japan this October, and the subsequent publication cited as reference (1).

## REFERENCES

1. Claybaugh, J.R., and M.R. Eichinger. Effects of Hypoxia on the Physiological Responses to Hypotensive Stress. In. *The 1997 Nagano Symposium on Sports Sciences*. H. Nose, T. Morimoto, and E.R. Nadel (eds.) Cooper Publishing Group, Carmel, IN., 1998, Chapter 28, pp 209-221.
2. Peuler, J.D., P.G. Schmid, D.A. Morgan, and A. Mark. Inhibition of renal sympathetic activity and heart rate by vasopressin in hemorrhaged diabetes insipitus rats. Am. J. Physiol. 258 (Heart Circ. Physiol. 27): H706-H712, 1990.
3. Eichinger, M.R. and J.R. Claybaugh: Hypoxia attenuates the renin response to hemorrhage. Am J Physiol 263 (Regulatory Integrative Comp. Physiol. 32): R664 - R669, 1992.
4. Resta, T.C., R.D. Russ, M.P. Doyle, J.M. Martinez, B.R. Walker. Cardiovascular responses to hemorrhage during acute and chronic hypoxia. Am. J. Physiol. 267 (Regulatory Integrative Comp. Physiol. 36): R619-R627, 1994.
5. Resta, T.C., J.M. Resta, and B.R. Walker. Role of endogenous opioids and serotonin in the hemodynamic response to hemorrhage during hypoxia. Am. J. Physiol. 269 (Heart, Circ. Physiol. 38): H1597-H1606, 1995.
6. Share, L. and M.N. Levy. Effect of carotid chemoreceptor stimulation on plasma antidiuretic hormone titer. Am. J. Physiol. 210:157-161, 1966.

7. Alexander, D.P., M.L. Forsling, M.J. Martin, D.A. Nixon, J.G. Ratcliff, D. Redstone, and D. Turnbridge. The effect of maternal hypoxia on fetal pituitary hormone release in the sheep. *Biol. Neonate.* 21:219-228, 1972.
8. Claybaugh, J.R. Wade, C.E., and Cucinell, S.A.: Fluid and electrolyte balance and hormonal responses to the hypoxic environment. In: *Hormonal Regulation of fluid and electrolytes: Environmental effects* (eds) J.R. Claybaugh and C.E. Wade, Plenum Publishing Corp., New York, NY, 187-214, 1989
9. Uyehara, C.F.T., J.R. Claybaugh, L.S. Matsuda, A.K. Sato, and S.C. Van Scoy. Vasopressin (VP) improves blood gas exchange in the neonatal piglets via V1-receptor stimulation. *FASEB J.* 7:A434 (Abstract 2512), 1993.
10. VanScoy, S.C., C.F.T. Uyehara, A.K. Sato, J.R. Claybaugh, and K.T. Nakamura. Improved gas exchange with vasopressin V1-agonist (V1) in newborn piglets is not seen with other vasoactive agents. *Experimental Biology* 94. Abstract # 4780, *FASEB J.* 8:A824, 1994.
11. Claybaugh, J.R. and C.F.T. Uyehara. Metabolism of Neurohypophseal Hormones. In: *The Neurohypophysis: A Window on Brain Function. Annals of the New York Academy of Science* Vol. 689, Eds. W.G. North, A.M. Moses, and L.Share, New York Academy of Science, New York, 250-268, 1993.
12. Wade, C.E. and J.P. Hannon. Confounding factors in the hemorrhage of conscious swine: a retrospective study of physical restraint, splenectomy, and hyperthermia. *Circulatory Shock* 24:175-182, 1988.
13. Horton, J.W., J.C. Longhurst, D. Coln, and J.H. Mitchell. Cardiovascular effects of hemorrhagic shock in spleen intact and in splenectomized dogs. *Clin. Physiol.* 4:533-548, 1984.
14. Budzikowski, A.S., P. Paczwa, and E. Szcepanska-Sadowska. Central V1 AVP receptors are involved in cardiovascular adaptation to hypovolemia in WKY but not SHR. *Am. J. Physiol.* 271 (Heart Circ. Physiol.40): H1057-H1064, 1996.
15. Magness, R.R., C.R. Parker Jr., and C.R. Rosenfeld. Systemic and uterine responses to chronic infusion of estradiol-17 $\beta$ . *Am. J. Physiol.* 265:E690-E698, 1993.
16. Crofton, J.T., L. Share, and D.P. Brooks. Pressor responsiveness to and secretion of vasopressin during states of the estrous cycle. *Am. J. Physiol.* 255:R1041-R1048, 1988.
17. Crofton, J.T., and L. Share. Sexual dimorphism in vasopressin and cardiovascular responses to hemorrhage in the rat. *Circulation Research* 66:1345-1353, 1990.
18. Banks, R.E., J.A. Davis, N.M. Coulson, and R.J. Beattie. A paracostal approach for splenectomy in the sheep. *J. Investigative Surg.* 1:143-148, 1988.
19. Atkins, J., K. Johnson, and F. Pearce. Oxygen inhalation after hemorrhage increases mean arterial blood pressure (MABP) even when changes in PaCO<sub>2</sub> are prevented. *FASEB J.* 10:A598 (Abstract 3450), 1996.
20. Uyehara, C.F.T., Hashiro, G.M., Lee, W.Q., Whippo, P.E., and Claybaugh, J.R., Vasopressin improves blood gases in the hypoxia-induced pulmonary hypertension via regional blood flow redistribution. *Experimental Biology '97*. New Orleans, LA. April 6-10, 1997. *FASEB J.* 11:A462, 1997 (Abstract #2674)